REVIEW ARTICLE

Multiple thermoregulatory effectors with independent central controls

Robin M. McAllen · Mutsumi Tanaka · Yoichiro Ootsuka · Michael J. McKinley

Accepted: 9 November 2009 / Published online: 1 December 2009 © Springer-Verlag 2009

Abstract This review first considers how mammalian body temperature regulation evolved, and how the brain's responses to thermoregulatory challenges are likely to be organised differently from the way an engineer would design them. This is because thermoregulatory effector mechanisms would have evolved one at a time, with each being superimposed on pre-existing mechanisms. There may be no functional need for the final ensemble of control loops to be coordinated by neural cross-connections: appropriate thermal thresholds would solve the problem sufficiently. Investigations first into thermoregulatory behaviours and later into unconscious thermoregulatory mechanisms (autonomic and shivering) have led investigators to the realisation that multiple control loops exist in the brain, with each effector system apparently regulated by its own central temperature sensors. This theme is developed with reference to data on four temperature-regulated neural outflows that have been studied on anaesthetized rats under standard conditions in the authors' laboratory. Direct comparisons were

Communicated by Osamu Shido.

R. M. McAllen (&) · M. Tanaka · Y. Ootsuka · M. J. McKinley Howard Florey Institute, University of Melbourne, Melbourne, VIC 3010, Australia e-mail: rmca@florey.edu.au

R. M. McAllen Department of Anatomy and Cell Biology, University of Melbourne, Melbourne, VIC 3010, Australia

Y. Ootsuka School of Medicine, Flinders University, Adelaide, SA 5042, Australia

M. J. McKinley Department of Physiology, University of Melbourne, Melbourne, VIC 3010, Australia

made between the behaviour of sympathetic nerves supplying the tail vasculature, vessels in the proximal hairy skin, interscapular brown adipose tissue (BAT) and fusimotor fibres to hind limb muscle. All four outflows were activated by cooling the skin, and all were silenced by neuronal inhibition in the medullary raphé. Their thermal thresholds were quite different, however, as were their relative responsiveness to core temperature. This was ranked as: tail > back skin > BAT > fusimotor. These and other data indicate that the four thermoeffector outflows are driven by separate neural pathways, each regulated by independent brain temperature sensors.

Keywords Rat tail · Cutaneous vasomotor · BAT · Fusimotor · Preoptic · Thermoregulation

Evolution of central temperature control in mammals

A number of mechanisms have evolved in mammals to regulate their body temperature, and different species may handle the same challenge in different ways. Evaporative cooling to lose heat in a hot environment, for example, occurs mainly by panting in dogs, by sweating in humans and by salivation with grooming in rats. On the colddefence side, the relative importance of shivering versus non-shivering thermogenesis also varies across species. In humans and dogs, for example, shivering is the predominant mechanism, whilst in rodents, non-shivering thermogenesis by brown adipose tissue (BAT) is prominent. Although the presence of BAT in adult humans has been doubted, recent evidence clearly shows that it is present and activated by cold (Zingaretti et al. [2009;](#page-6-0) Saito et al. [2009;](#page-6-1) van Marken Lichtenbelt et al. [2009;](#page-6-2) Cypess et al. [2009](#page-5-0)). In all mammals, cutaneous vasoconstriction reduces heat loss.

Considering these matters, it has been noted that we should not expect evolution to generate natural control systems that operate exactly as an engineer would design them (Partridge [1982\)](#page-5-1). All that is required for a new mechanism to be conserved is that it should improve the animal's chance of survival and reproduction; it does not matter how elegantly the physiological problem is solved (Partridge [1982](#page-5-1); Gould 1983). The low occurrence rate of beneficial mutations results in new features being added one at a time, with each being superimposed on previous adaptations (Gould [1983](#page-5-2)). In the evolution of cold defences, for example, non-shivering thermogenesis by BAT is a mammalian 'invention' (Cannon and Nedergaard [2004](#page-5-3)) that would have been superimposed on pre-existing ancestral responses such as warm-seeking behaviour (Seebacher and Franklin [2005](#page-6-3)). Our reptilian ancestors may well have been able to control blood flow to their skin: modern reptiles such as crocodiles (Seebacher and Franklin [2005\)](#page-6-3) and iguanas (Morgareidge and White [1969](#page-5-4)) are able to vasodilate their skin when basking in the warmth and to vasoconstrict in the cold. Those responses, however, appear to be mediated by local temperature rather than by vasomotor nerves (Morgareidge and White [1969](#page-5-4)). Whilst human cutaneous vessels also respond directly to temperature, dilating with heat and constricting with cold, their behaviour is generally dominated by neural drive from the CNS (Johnson and Proppe [1996\)](#page-5-5). Successive evolutionary innovations may thus address a common challenge (in this case, defence against cold), and each will be conserved if its action is independently beneficial. As argued by Partridge, there may be no need for such mechanisms to be coordinated by common neural circuitry: only if neural coordination confers a further significant advantage is it likely to evolve (Partridge [1982](#page-5-1)).

Multiple control mechanisms

It has been known for some time that thermoregulatory behaviours are organised by the brain independently of autonomic mechanisms, and indeed survive brain lesions of the preoptic area that disable the latter (Satinoff and Rutstein [1970](#page-6-4); Almeida et al. [2006](#page-5-6)). Moreover, it became evident that distinct pathways *with anatomically distinct central thermosensors* regulate different heat-defence behaviours, such as locomotion, saliva spreading and postural extension in rats (Roberts [1988](#page-5-7)). Evidence also shows that involuntary control mechanisms (i.e. autonomic and shivering) are multiple and parallel rather than unitary. In 1978, Gilbert and Blatteis showed that preoptic knife cuts could disable cutaneous vasoconstriction, but shivering was left unaffected (Gilbert and Blatteis [1977\)](#page-5-8). In the 1990s, Kanosue et al. pointed to a series of differences in the

descending pathways from preoptic warm-sensitive neu-rons to different effectors (Kanosue et al. [1994a](#page-5-9), [1994b;](#page-5-10) Chen et al. [1998;](#page-5-11) Zhang et al. [1995\)](#page-6-5). The logical inference was that, instead of a single central 'thermostat' regulating a suite of heat loss and heat gain mechanisms, the preoptic region contains a number of independent groups of temperature-sensitive neurons, each regulating individual effector mechanisms (Nagashima et al. [2000](#page-5-12); Romanovsky [2007](#page-6-6)). The present study extends this theme of multiple independent controllers operating via distinct efferent pathways. Regardless of whether this principle applies universally to mammalian body temperature control, it is clear that one cannot transfer lessons learnt from one thermoeffector control system directly to another.

The principal difficulty with this view is that the mechanisms regulating body temperature appear to act seamlessly together as a precisely coordinated control system seeking a 'set point'. This applies not only to the basal state, but parallel threshold shifts can cause a regulated 'resetting' of body temperature upwards or downwards with (e.g.) diurnal body temperature variation, fever and hypoxia (Cabanac [2006\)](#page-5-13). On some occasions, however, this coordination can break down: for example, high doses of endotoxin cause a regulated hypothermia that involves cold-seeking behaviour and a substantially reduced temperature threshold for thermogenesis, but only a minimal change in cutaneous vasoconstrictor threshold (Almeida et al. [2006](#page-5-6); Romanovsky et al. [1996\)](#page-6-7). Divergence of thresholds between different thermoeffector responses has also been noted in the later phases of fever (Vybiral et al. [1987](#page-6-8)). Our view is that the occurrence of parallel threshold shifts between the various thermoeffectors is a beguiling phenomenon that requires further explanation. It does not amount to compelling evidence for the control of multiple effectors by a single brain centre.

The medullary raphé

From the late 1990s, it became apparent from the work in several laboratories that a critical role in cold defence was played by neurons in a small region of the ventral medullary raphé, at the rostrocaudal level of the caudal part of the facial nucleus (Rathner and McAllen [1999;](#page-5-14) Blessing et al. [1999](#page-5-15); Morrison et al. [1999](#page-5-16); Morrison [1999](#page-5-17); Rathner et al. [2001](#page-5-18); Tanaka et al. [2002](#page-6-9)). Inhibition of neurons in this small brainstem region could suppress both non-shivering thermogenesis by BAT and cutaneous vasoconstriction in response to cold exposure or experimental fever (Tanaka et al. [2002](#page-6-9); Nakamura et al. [2002;](#page-5-19) Morrison [2003](#page-5-20); Madden and Morrison [2003](#page-5-21); Nakamura and Morrison [2007;](#page-5-22) Korsak and Gilbey [2004;](#page-5-23) Ootsuka et al. [2004;](#page-5-24) Rathner et al. [2008;](#page-5-25) Tanaka et al. [2007\)](#page-6-10). In conscious rats, inactivating neurons

of this raphe region rendered the animals unable to defend their body temperature against cold [\(Zaretsky et al. 2003](#page-6-11)). Neurons in this same region had previously been implicated as mediating the gastric secretory and motility responses characteristic of rats exposed to cold (Tache et al. [1995](#page-6-12); Yang et al. [2000\)](#page-6-13). Could there be, after all, a coordinating centre in this discrete brainstem region for multiple colddefence responses? Others (Rathner et al. [2008](#page-5-25)) and we (Tanaka et al. [2006,](#page-6-14) [2007](#page-6-10); Ootsuka and McAllen [2006\)](#page-5-26) investigated this possibility by directly comparing several cold-defence effector pathways in anaesthetized rats.

Studies comparing four cold-defence effector pathways

The studies from our laboratory have been performed on animals under standard experimental conditions. Male Sprague–Dawley rats were anaesthetized with urethane $(1-1.5 \text{ g/kg}$ i.v.) after preparatory surgery under 2% isofluorane; they were ventilated artificially with oxygen via a tracheostomy. The animal's head was fixed in a stereotaxic frame and its shaved trunk was encased in a water-perfused silastic jacket. Body core temperature was measured from the rectum and skin temperature as the mean value of thermocouples placed at three sites under the water jacket. No paralysing agent was given. Nerve activity was recorded from peripheral thermoeffector nerves, usually in pairs. The standard cooling protocol started from warm conditions. Cold water was then pumped through the water jacket for successive brief (0.5–2 min) episodes that were separated by similar time intervals, during which warm water was reperfused. Core temperature fell in response to the repeated skin cooling episodes. The different time courses of skin and core temperature falls allowed the neural response to each to be distinguished and measured (Owens et al. [2002](#page-5-27)).

Whilst the use of general anaesthesia is a limitation, the use of parallel neural recordings and standard experimental conditions is an aid to interpreting the findings.

Case 1: tail vasoconstrictior and BAT thermogenic nerve activity

Recordings of sympathetic nerves supplying the tail vasculature and interscapular BAT have revealed similarities and differences. Both nerves show little or no activity when skin and core were kept at warm temperatures (Johnson and Gilbey [1998](#page-5-28); Owens et al. [2002;](#page-5-27) Morrison [2003;](#page-5-20) Ootsuka et al. [2004](#page-5-24); Nakamura and Morrison [2008a](#page-5-29)). Both nerves may be activated by cooling and during experimental fever after prostaglandin E_2 (or E_1) injection into the brain (Johnson and Gilbey [1998](#page-5-28); Owens et al. [2002;](#page-5-27) Morrison [2003](#page-5-20); Collins et al. [2002;](#page-5-30) Ootsuka et al. [2004;](#page-5-24) Nakamura and Morrison [2008a;](#page-5-29) Tanaka and McAllen [2005\)](#page-6-15). Tonic and evoked activity in both nerves may be eliminated by inhibiting neurons in the ventral medullary raphé at the level of the caudal part of the facial nucleus (Korsak and Gilbey [2004;](#page-5-23) Morrison [2003;](#page-5-20) Ootsuka et al. [2004;](#page-5-24) Nakamura and Morrison [2008a](#page-5-29)).

When their activity was compared in simultaneous recordings (Fig. [1a](#page-3-0)), both nerves were activated by cooling the skin of the trunk, but this occurred at different threshold temperatures (Fig. [1](#page-3-0)a). Further repeated skin cooling episodes caused the core temperature to fall, whereupon tail fibre activity started responding to this signal. As core temperature fell further, tail fibres were no longer silenced each time the skin was rewarmed. BAT nerve activity, however, continued to respond only to skin temperature until core temperature fell substantially further (Fig. [1a](#page-3-0)). The relative sensitivity of BAT nerve activity to core temperature was always much less than that of tail fibre activity, and in two out of seven cases examined in that study, no component of BAT nerve activity was attributable to core cooling (Ootsuka and McAllen [2006](#page-5-26)).

It is thus clear that the central mechanisms driving these two sympathetic outflows cannot be identical. Further corroboration of this point is provided by the lack of coherence between the frequency spectra of the two nerves' activity, other than a component associated with the ventilatory cycle (Ootsuka and McAllen [2006\)](#page-5-26). Moreover, differences have been found between their central pathways. Tail fibre activity is supported by an auxiliary, excitatory drive from neurons in the rostral ventrolateral medulla (RVLM) (Ootsuka and McAllen [2005\)](#page-5-31); BAT nerve activity, by contrast, is inhibited by RVLM neurons (Morrison [1999](#page-5-17)). Most strikingly, it has recently been shown that whilst BAT nerve activity depends on a critical synapse in the dorsomedial hypothalamus (DMH), tail activity is unaffected by inhibition of DMH neurons (Rathner et al. [2008\)](#page-5-25).

Case 2: tail and back skin cutaneous vasoconstrictor fibre activity

In humans, differences have been documented between the neural control of blood flow to the acral versus proximal skin regions (Johnson and Proppe [1996](#page-5-5)). In rats, the cutaneous vasomotor drives to the hind paw and tail are very similar (Smith and Gilbey [2000\)](#page-6-16). Only one study has directly compared the neural activity supplying these distal sites with that to more proximal skin regions (Tanaka et al. [2007](#page-6-10)). Figure [1](#page-3-0)b is taken from that study and shows how the cutaneous vasoconstrictor (CVC) activity to the proximal hairy skin of the back has different thresholds for activation by both skin and core cooling. Moreover, the ratio of responsiveness to core versus skin temperature was quite

Fig. 1 Representative chart recordings of sympathetic nerve activity (SNA) supplying several thermoregulatory effector organs in the rat showing responses to repeated brief episodes of skin cooling, whilst core temperature was allowed to fall. **a** Comparison of SNA supplying brown adipose tissue (BAT) with that supplying tail vessels. Note the different thresholds to skin and core cooling, as well as the much stronger response to core cooling by tail than by BAT SNA (Ootsuka and McAllen [2006\)](#page-5-26). **b** Comparison of SNA supplying back skin versus that to tail vessels. Note the different thresholds and the lesser response to

core cooling by back skin SNA compared with tail SNA (Tanaka et al. [2007](#page-6-10)). **c** Fusimotor fibre activity supplying the gastrocnemius muscle. Note how these fibres are activated by skin cooling, but not by core cooling, even when it fell to 33°C (From reference Tanaka et al. [2006](#page-6-14)). **d** Comparison of SNA supplying BAT and back skin. Note that the back skin SNA is activated earlier by skin cooling, and shows a proportionately greater response to core temperature than does BAT SNA (MT, MMcK and RMcA, unpublished data)

different between proximal CVC and tail fibre activity (Fig. [1b](#page-3-0)). Finally, there was a lack of coherence between the activity spectra of the two nerves in the frequency domain (Tanaka et al. [2007\)](#page-6-10).

Although both back skin CVC and tail fibre activity may be silenced by inhibiting raphé neurons (Tanaka et al. [2007\)](#page-6-10), it appears that their central drive pathways must be different.

Case 3: fusimotor fibre activity

A further 'unconscious' cold-defence response was recently reported: activation of fusimotor fibres to the limb muscles (Tanaka et al. 2006). The significance of this response may be twofold: first, it could underlie the increased muscle tone that precedes overt shivering in the cold (Meigal et al. [2003\)](#page-5-32); second, it would enhance the stretch reflex, on which shivering is heavily dependent (Perkins [1945\)](#page-5-33) and may thereby facilitate shivering.

As shown in Fig. $1c$ $1c$, fusimotor fibres were silent in anaesthetized rats until activated by skin cooling. The remarkable feature that we wish to emphasise is that whilst repeated skin cooling episodes reliably activated fusimotor fibres, they always fell silent when the skin was rewarmed; no component of activity was attributable to core cooling, even when core temperature fell as low as 33°C (Fig. [1c](#page-3-0)). There was, however, evidence for a minor inhibitory action of warm core temperature on the fusimotor response to skin cooling (Tanaka et al. 2006). This effect may be mediated by preoptic temperature-sensitive neurons (Von Euler and Soderberg [1957](#page-6-17)). Like BAT and tail and back skin CVC fibre activity, fusimotor activation by cold skin was abolished by inhibiting medullary raphé neurons (Tanaka et al. [2006](#page-6-14)).

Case 4: comparison of BAT and back skin CVC activity

Both BAT and back skin CVC activity occupy an intermediate position between tail and fusimotor fibres, with respect to their thermal thresholds and their relative responsiveness to skin versus core temperature (Fig. [2a](#page-4-0)). Their

Fig. 2 a The relative responsiveness to skin and core temperatures of four cold-activated neural outflows: tail vessels, back skin vessels, brown adipose tissue (BAT) and fusimotor fibres. **b** Inferred neural pathways driving those four neural outflows. The hypothesis is that each is driven by independent neural pathways regulated by distinct preoptic neurons with different intrinsic thermosensitivity (indicated by the length of the thermometer shaft)

preganglionic axons both emerge from a similar level of the thoracic spinal cord. Could they perhaps be receiving copies of the same neural drive signals? To answer this question, simultaneous recordings from the two nerves were made in two rats (M. Tanaka, M.J. McKinley and R.M. McAllen, unpublished data). As may be seen from the record excerpt shown in Fig. [1](#page-3-0)d, their response patterns were distinct. Back skin CVC fibres were activated earlier during skin cooling and showed a greater relative response to core temperature than did BAT nerve activity.

Discussion

Our hypothesis for how these four neural outflows receive their temperature-sensitive drives is illustrated diagrammatically in Fig. [2](#page-4-0)b. The key point (cf. Nagashima et al. [2000\)](#page-5-12) is that these efferent pathways are entirely separate from the preoptic area downwards. Whilst it has not been formally shown that back skin CVC and fusimotor activity are regulated by preoptic neurons, we consider it most probable that they are. A further unknown point is whether back skin CVC or fusimotor pathways involve a relay in the dorsomedial hypothalamus (DMH), or indeed elsewhere.

What we can infer is that the preoptic neurons regulating tail fibre activity must be much more sensitive to core temperature than those regulating other outflows. In contrast, the neurons regulating the fusimotor outflow evidently possess minimal intrinsic core temperature sensitivity (the

relative sensitivity to core temperature in each pathway is represented in Fig. [2b](#page-4-0) by the length of the thermometer shaft). It seems likely that similar afferent pathways, via lamina 1 of the spinal cord dorsal horn and the parabrachial nuclei, convey skin temperature signals to all of these effector mechanisms (Nakamura and Morrison [2008a\)](#page-5-29). Again, it is assumed by analogy with the BAT pathway (Nakamura and Morrison [2008a](#page-5-29); Osaka [2004](#page-5-34); Nakamura and Morrison [2008b](#page-5-35)) that the main input from skin temperature signals to each of these other cold-defence pathways is relayed via preoptic neurons. These points await direct confirmation.

The significance of the obligatory synaptic relay in the medullary raphé for all four pathways considered here, as well as others (Tache et al. [1995;](#page-6-12) [Zaretsky et al. 2003](#page-6-18)), remains an enigma. The idea that this synapse might be a subsidiary site, where humoral factors such as prostaglan- $\sin E_2$ might act on several pathways at once to cause fever. received no experimental support (Tanaka and McAllen [2005](#page-6-15)). A further point that deserves consideration is that the abundant serotonergic neurons in this region, whilst they may not constitute the primary drive pathway for thermogenesis or vasoconstriction (Nakamura et al. [2004](#page-5-36)), nevertheless, could play a modulatory role that reinforces several cold-defence responses at a spinal level (Madden and Morrison [2008;](#page-5-37) Marina et al. [2006\)](#page-5-38). Serotonergic neurons in this region are activated by cold exposure (Yang et al. [2000;](#page-6-13) Nakamura et al. [2004\)](#page-5-36) and may receive their temperature-related signals directly or indirectly from the preoptic area. Alternatively or additionally, they might receive signals from cold skin by more direct pathways that are contained within the midbrain and brain stem. This possibility should be considered because in decerebrate rats, cold exposure is still able to increase BAT norepinephrine turnover (indicating sympathetic activation) (Nautiyal et al. [2008](#page-5-39)) and neurons responding to skin temperature may still be recorded in the medullary raphé (Dickenson [1977\)](#page-5-40).

Conclusions

The four thermoeffector outflows considered here are regulated by parallel efferent pathways; they all relay synaptically in the medullary raphé, but do so without any apparent neural cross talk. We infer that distinct preoptic neurons with differing intrinsic temperature sensitivity regulate each effector. The task remains to link individual effectors with their own central thermoregulatory control neurons.

Acknowledgments RMcA and MMcK hold NHMRC Fellowships (566667 and 454369). YO held a fellowship from the High Blood Pressure Research Council of Australia. The work was supported by NHM-RC project grants 232305 and 454601, the Robert J Jr and Helen C Kleberg Foundation and the G Harold and Leila Y Mathers Foundation.

References

- Almeida MC, Steiner AA, Branco LG, Romanovsky AA (2006) Neural substrate of cold-seeking behavior in endotoxin shock. PLoS One 1:e1
- Blessing WW, Yu YH, Nalivaiko E (1999) Raphe pallidus and parapyramidal neurons regulate ear pinna vascular conductance in the rabbit. Neurosci Lett 270(1):33–36
- Cabanac M (2006) Adjustable set point: to honor Harold T. Hammel. J Appl Physiol 100(4):1338–1346
- Cannon B, Nedergaard J (2004) Brown adipose tissue: function and physiological significance. Physiol Rev 84(1):277–359
- Chen XM, Hosono T, Yoda T, Fukuda Y, Kanosue K (1998) Efferent projection from the preoptic area for the control of non-shivering thermogenesis in rats. J Physiol 512(Pt 3):883– 892
- Collins DR, Korsak A, Gilbey MP (2002) Cutaneous sympathetic motor rhythms during a fever-like response induced by prostaglandin E(1). Neuroscience 110(2):351–360
- Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB et al (2009) Identification and importance of brown adipose tissue in adult humans. N Engl J Med 360(15):1509–1517
- Dickenson AH (1977) Specific responses of rat raphe neurones to skin temperature. J Physiol 273(1):277–293
- Gilbert TM, Blatteis CM (1977) Hypothalamic thermoregulatory pathways in the rat. J Appl Physiol 43(5):770–777
- Gould S (1983) The panda's thumb: more reflections on natural history. Penguin Books, Harmondsworth
- Johnson CD, Gilbey MP (1998) Focally recorded single sympathetic postganglionic neuronal activity supplying rat lateral tail vein. J Physiol 508(Pt 2):575–585
- Johnson JM, Proppe DW (1996) Cardiovascular adjustments to heat stress. In: Fregley M, Blatteis C (eds) Handbook of physiology: section 4, environmental physiology. American Physiological Society, Washington, DC
- Kanosue K, Zhang YH, Yanase-Fujiwara M, Hosono T (1994a) Hypothalamic network for thermoregulatory shivering. Am J Physiol 267(1 Pt 2):R275–R282
- Kanosue K, Yanase-Fujiwara M, Hosono T (1994b) Hypothalamic network for thermoregulatory vasomotor control. Am J Physiol 267(1 Pt 2):R283–R288
- Korsak A, Gilbey MP (2004) Rostral ventromedial medulla and the control of cutaneous vasoconstrictor activity following i.c.v. prostaglandin E(1). Neuroscience 124(3):709–717
- Madden CJ, Morrison SF (2003) Excitatory amino acid receptor activation in the raphe pallidus area mediates prostaglandin-evoked thermogenesis. Neuroscience 122(1):5–15
- Madden CJ, Morrison SF (2008) Brown adipose tissue sympathetic nerve activity is potentiated by activation of 5-hydroxytryptamine (5-HT)1A/5-HT7 receptors in the rat spinal cord. Neuropharmacology 54(3):487–496
- Marina N, Taheri M, Gilbey MP (2006) Generation of a physiological sympathetic motor rhythm in the rat following spinal application of 5-HT. J Physiol 571(Pt 2):441–450
- Meigal AY, Oksa J, Gerasimova LI, Hohtola E, Lupandin YV, Rintamaki H (2003) Force control of isometric elbow flexion with visual feedback in cold with and without shivering. Aviat Space Environ Med 74(8):816–821
- Morgareidge KR, White FN (1969) Cutaneous vascular changes during heating and cooling in the Galapagos marine iguana. Nature 223(5206):587–591
- Morrison SF (1999) RVLM and raphe differentially regulate sympathetic outflows to splanchnic and brown adipose tissue. Am J Physiol 276(4 Pt 2):R962–R973
- Morrison SF (2003) Raphe pallidus neurons mediate prostaglandin E2-evoked increases in brown adipose tissue thermogenesis. Neuroscience 121(1):17–24
- Morrison SF, Sved AF, Passerin AM (1999) GABA-mediated inhibition of raphe pallidus neurons regulates sympathetic outflow to brown adipose tissue. Am J Physiol 276(2 Pt 2):R290–R297
- Nagashima K, Nakai S, Tanaka M, Kanosue K (2000) Neuronal circuitries involved in thermoregulation. Auton Neurosci 85(1–3):18–25
- Nakamura K, Morrison SF (2007) Central efferent pathways mediating skin cooling-evoked sympathetic thermogenesis in brown adipose tissue. Am J Physiol Regul Integr Comp Physiol 292(1):R127– R136
- Nakamura K, Morrison SF (2008a) A thermosensory pathway that controls body temperature. Nat Neurosci 11(1):62–71
- Nakamura K, Morrison SF (2008b) Preoptic mechanism for colddefensive responses to skin cooling. J Physiol 586(10):2611– 2620
- Nakamura K, Matsumura K, Kaneko T, Kobayashi S, Katoh H, Negishi M (2002) The rostral raphe pallidus nucleus mediates pyrogenic transmission from the preoptic area. J Neurosci 22(11):4600–4610
- Nakamura K, Matsumura K, Hubschle T, Nakamura Y, Hioki H, Fujiyama F et al (2004) Identification of sympathetic premotor neurons in medullary raphe regions mediating fever and other thermoregulatory functions. J Neurosci 24(23):5370–5380
- Nautiyal KM, Dailey M, Brito N, Brito MN, Harris RB, Bartness TJ et al (2008) Energetic responses to cold temperatures in rats lacking forebrain–caudal brain stem connections. Am J Physiol Regul Integr Comp Physiol 295(3):R789–R798
- Ootsuka Y, McAllen RM (2005) Interactive drives from two brain stem premotor nuclei are essential to support rat tail sympathetic activity. Am J Physiol Regul Integr Comp Physiol 289(4):R1107– R1115
- Ootsuka Y, McAllen RM (2006) Comparison between two rat sympathetic pathways activated in cold defense. Am J Physiol Regul Integr Comp Physiol 291(3):R589–R595
- Ootsuka Y, Blessing WW, McAllen RM (2004) Inhibition of rostral medullary raphe neurons prevents cold-induced activity in sympathetic nerves to rat tail and rabbit ear arteries. Neurosci Lett 357(1):58–62
- Osaka T (2004) Cold-induced thermogenesis mediated by GABA in the preoptic area of anesthetized rats. Am J Physiol Regul Integr Comp Physiol 287(2):R306–R313
- Owens NC, Ootsuka Y, Kanosue K, McAllen RM (2002) Thermoregulatory control of sympathetic fibres supplying the rat's tail. J Physiol 543(Pt 3):849–858
- Partridge LD (1982) The good enough calculi of evolving control systems: evolution is not engineering. Am J Physiol 242(3):R173– R177
- Perkins JF Jr (1945) The role of proprioceptors in shivering. Am J Physiol 145(2):264–271
- Rathner JA, McAllen RM (1999) Differential control of sympathetic drive to the rat tail artery and kidney by medullary premotor cell groups. Brain Res 834(1–2):196–199
- Rathner JA, Owens NC, McAllen RM (2001) Cold-activated raphespinal neurons in rats. J Physiol 535(Pt 3):841–854
- Rathner JA, Madden CJ, Morrison SF (2008) Central pathway for spontaneous and prostaglandin E2-evoked cutaneous vasoconstriction. Am J Physiol Regul Integr Comp Physiol 295(1):R343– R354
- Roberts WW (1988) Differential thermosensor control of thermoregulatory grooming, locomotion, and relaxed postural extension. Ann NY Acad Sci 525:363–374
- Romanovsky AA (2007) Thermoregulation: some concepts have changed. Functional architecture of the thermoregulatory system. Am J Physiol Regul Integr Comp Physiol 292(1):R37–R46
- Romanovsky AA, Shido O, Sakurada S, Sugimoto N, Nagasaka T (1996) Endotoxin shock: thermoregulatory mechanisms. Am J Physiol 270(4 Pt 2):R693–R703
- Saito M, Okamatsu-Ogura Y, Matsushita M, Watanabe K, Yoneshiro T, Nio-Kobayashi J et al (2009) High incidence of metabolically active brown adipose tissue in healthy adult humans: effects of cold exposure and adiposity. Diabetes 58(7):1526–1531
- Satinoff E, Rutstein J (1970) Behavioral thermoregulation in rats with anterior hypothalamic lesions. J Comp Physiol Psychol 71(1):77–82
- Seebacher F, Franklin CE (2005) Physiological mechanisms of thermoregulation in reptiles: a review. J Comp Physiol B 175(8):533–541
- Smith JE, Gilbey MP (2000) Coherent rhythmic discharges in sympathetic nerves supplying thermoregulatory circulations in the rat. J Physiol 523(Pt 2):449–457
- Tache Y, Yang H, Kaneko H (1995) Caudal raphe-dorsal vagal complex peptidergic projections: role in gastric vagal control. Peptides 16(3):431–435
- Tanaka M, McAllen RM (2005) A subsidiary fever center in the medullary raphe? Am J Physiol Regul Integr Comp Physiol 289(6):R1592–R1598
- Tanaka M, Nagashima K, McAllen RM, Kanosue K (2002) Role of the medullary raphe in thermoregulatory vasomotor control in rats. J Physiol 540(Pt 2):657–664
- Tanaka M, Owens NC, Nagashima K, Kanosue K, McAllen RM (2006) Reflex activation of rat fusimotor neurons by body surface cooling, and its dependence on the medullary raphe. J Physiol 572(Pt 2):569–583
- Tanaka M, Ootsuka Y, McKinley MJ, McAllen RM (2007) Independent vasomotor control of rat tail and proximal hairy skin. J Physiol 582(Pt 1):421–433
- van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND et al (2009) Cold-activated brown adipose tissue in healthy men. N Engl J Med 360(15):1500–1508
- Von Euler C, Soderberg U (1957) The influence of hypothalamic thermoceptive structures on the electroencephalogram and gamma motor activity. Electroencephalogr Clin Neurophysiol 9(3):391–408
- Vybiral S, Szekely M, Jansky L, Cerny L (1987) Thermoregulation of the rabbit during the late phase of endotoxin fever. Pflugers Arch 410(1–2):220–222
- Yang H, Yuan PQ, Wang L, Tache Y (2000) Activation of the parapyramidal region in the ventral medulla stimulates gastric acid secretion through vagal pathways in rats. Neuroscience 95(3):773–779
- Zaretsky DV, Zaretskaia MV, DiMicco JA (2003a) Stimulation and blockade of $GABA(A)$ receptors in the raphe pallidus: effects on body temperature, heart rate, and blood pressure in conscious rats. Am J Physiol Regul Integr Comp Physiol 285(1):R110–R116
- Zaretsky DV, Zaretskaia MV, Samuels BC, Cluxton LK, DiMicco JA (2003b) Microinjection of muscimol into raphe pallidus suppresses tachycardia associated with air stress in conscious rats. J Physiol 546(Pt 1):243–250
- Zhang YH, Yanase-Fujiwara M, Hosono T, Kanosue K (1995) Warm and cold signals from the preoptic area: which contribute more to the control of shivering in rats? J Physiol 485(Pt 1):195–202
- Zingaretti MC, Crosta F, Vitali A, Guerrieri M, Frontini A, Cannon B et al (2009) The presence of UCP1 demonstrates that metabolically active adipose tissue in the neck of adult humans truly represents brown adipose tissue. FASEB J 23(9):3113–3120